Vegetation Response to Timing of Discing to Manage Northern Bobwhite Habitat in Texas

Fidel Hernandez, Caesar Kleberg Wildlife Research Institute, Texas A&M University-Kingsville, 700 University Blvd, MSC 218, Kingsville, TX 78363-8202

J. Lane Roberson,¹ Caesar Kleberg Wildlife Research Institute, Texas A&M University-Kingsville, 700 University Blvd, MSC 218, Kingsville, TX 78363-8202

Ralph L. Bingham, Caesar Kleberg Wildlife Research Institute, Texas A&M University-Kingsville, 700 University Blvd, MSC 218, Kingsville, TX 78363-8202

Stephen J. DeMaso, Caesar Kleberg Wildlife Research Institute, Texas A&M University-Kingsville, 700 University Blvd, MSC 218, Kingsville, TX 78363-8202

Robert M. Perez, Texas Parks and Wildlife Department, La Vernia, TX 78121

Timothy E. Fulbright, *Caesar Kleberg Wildlife Research Institute*, *Texas A&M University-Kingsville*, 700 University Blvd, MSC 218, Kingsville, TX 78363-8202

Leonard A. Brennan, Caesar Kleberg Wildlife Research Institute, Texas A&M University-Kingsville, 700 University Blvd, MSC 218, Kingsville, TX 78363-8202

Abstract: Discing is commonly recommended to improve northern bobwhite (*Colinus virginianus*) habitat. However, little information exists regarding optimal timing of discing or the duration of discing effects on semiarid rangelands. Our objectives were to evaluate vegetation response to autumn (October 2003), winter (January 2004), and spring (March 2004) discing in two ecoregions of Texas (Rio Grande Plains and Rolling Plains). Our study design was a completely randomized, two-factor (treatment and soil texture) factorial with repeated measures. We collected data on percent bare ground, forb density, visual obstruction, and non-native grass density during pretreatment (September 2003) and six sampling periods post-treatment (March, May, and July 2004 and 2005). Differences in percent bare ground, forb density, and visual obstruction differed (P>0.05) among treatments only during the first year post-discing in all soil textures and ecoregions. The trend was for density of non-native grasses to increase (>60%) on both control and disced plots in the Rio Grande Plains. Density of non-native grasses in the Rolling Plains exhibited no (P<0.05) change. Our findings suggest the positive, structural effects of discing on bobwhite habitat on semiarid rangelands can be short-lived (<1.5 years) and unintended, negative habitat changes may result from an increase of non-native grasses if additional measures such as herbicide application are not taken.

Key words: Colinus virginianus, discing, non-native grasses, northern bobwhite, Texas

Proc. Annu. Conf. Southeast. Assoc. Fish and Wildl. Agencies 63:61-69

A basic principle of wildlife habitat management is that animal species are adapted to certain seral stages of ecological succession and that provision of the correct seral stage will improve resource utilization and subsequently enhance abundance (Bailey 1984, Patton 1992, Robinson and Bolen 2003). Northern bobwhites (*Colinus virginianus*) generally are considered an early successional species (Stoddard 1931, Jackson 1972), though seral stage affiliation may vary with site productivity (Spears et al. 1993). Early seral stages provide important brood-rearing and foraging habitat for bobwhites (Stoddard 1931, Rosene 1969, Jackson et al. 1987). Consequently, management practices that disturb the soil, such

as discing, have been recommended to manage bobwhite habitat (Stoddard 1931, Rosene 1969, Lehmann 1984, Guthery 1986).

Discing patches of rangeland can improve bobwhite habitat by increasing percentage of bare ground (Webb and Guthery 1983, Greenfield et al. 2002), stimulating growth of important food plants (Peoples et al. 1994), and creating plant structural diversity necessary for invertebrates (Shelton and Edwards 1983, Manley et al. 1994). In semiarid rangelands, the optimal time for discing and its lasting effects on habitat structure are relatively unknown (Fulbright 1999). Recently, the value of discing for improving wildlife habitat has been questioned because soil disturbance may favor establishment of non-native grasses (Johnson and Fulbright 2008) which can be detrimental to bobwhites (Kuvlesky et al. 2002). The

^{1.} Current address: Magueyitos Ranch, P.O. Box 478, Encinal, TX 78019

objectives of our study were to 1) compare bare ground exposure, forb density, and visual obstruction among autumn (October), winter (January), and spring (March) discing over two growing seasons, and 2) document the response of non-native grasses to soil disturbance.

Study Area

Our study was conducted in the Rio Grande Plains and Rolling Plains ecoregions of Texas (Gould 1975). Specific study areas were the James E. Daughtery Wildlife Management Area (WMA) (Live Oak and McMullen counties) and Gene Howe WMA (Hemphill County). The James E. Daughtery WMA study area is in the Rio Grande Plains ecoregion (Gould 1975). It is a 1,781-ha low-fenced multiple-use recreational area. Mean annual temperature was 21.7 C with a mean rainfall of 71.1 cm and a 289-day growing season. Rainfall during the study period exceeded the long-term mean in 2004 (99.0 cm) and approximated the long-term mean in 2005 (61.3 cm). Predominant brush species include honey mesquite (Prosopis glandulosa), huisache (A. smallii), blackbrush (A. rigidula), granjeno (Celtis pallida), and pricklypear cactus (Optuntia lindheimeri). Native grasses common to the area included silver bluestem (Bothriochloa laguroides), curly mesquite (Hilaria belangeri), and Arizona cottontop (Digitaria californica). Predominant forbs include croton (Croton spp.), sunflower (Helianthus annuus), dayflower (Commelina erecta), and partridge pea (Chamaecrista fasciculata). Three species of non-native grasses were found in our study area: King Ranch bluestem (B. ischaeum), Kleberg bluestem (Dicanthium annulatum), and bermudagrass (Cynodon dactylon).

The Gene Howe WMA is in the Rolling Plains ecoregion (Gould 1975). The WMA consists of 2,382 ha located along the Canadian River in Hemphill County. Mean maximum temperature was 35 C in July with a mean minimum of -5 C in January. Mean annual rainfall was 52.1 cm, and the growing season averages 204 days a year. Rainfall during the 2004 study period was well above the longterm mean and approximated in 2005 (55.4 cm) the long-term average. The Gene Howe WMA is characterized as a sand sagebrush (Artemesia filifolia)-mid-grass rangeland and eastern cottonwood (Populus deltoides)-tallgrass bottomland along the river. Vegetation consisted predominantly of sand sagebrush, sand plum (Prunus angustifolia), fragrant sumac (Rhus aromatica), eastern cottonwood, netleaf hackberry (Celtis reticulata), and black locust (Robinia pseudoacacia). Predominant grasses included little bluestem (Schizachyrium scoparium), sand dropseed (Sporobolus cryptandrus), blue grama (Bouteloua gracilis), big bluestem (A. gerardii), switchgrass (Panicum virgatum), indiangrass (Sorghastrum nutans), and alkali sacaton (Sporobolus airoides). Common forbs included western ragweed (Ambrosia psilostachya), silverleaf nightshade (Solanum

Vegetation Response to Discing in Texas Rangelands Hernàndez et al. 62

elaeagnifolium), common sunflower, and Texas croton (*C. texensis*). Three species of non-native grasses occurred on the study site: perennial ryegrass (*Lolium perenne*), Japanese brome (*Bromus japonicus*), and cheatgrass (*Bromus tectorum*).

Methods

Experimental Design

Our study design was a completely randomized, two-factor factorial with repeated measures. The two factors were discing treatment and soil texture. We evaluated two soil textures (i.e., sandy or clay) in each ecoregion. Each soil-delineated study plot consisted of 15 experimental units (10×100 m) that were separated by 10-m buffer strips. Our treatments involved one control (no discing) and three seasons of discing: autumn (October 2003), winter (January 2004), and spring (March 2004). We randomly selected three experimental units within each soil texture for each discing treatment and six experimental units for control. Thus, each discing treatment had three replications within each soil-delineated plot, and the control had six replications (3 discing treatments \times 3 experimental units/treatment=9 experimental units; 1 control \times 6 experimental units/control=6 experimental units; for a total of 15 total experimental units per soil study plot). Treatment plots were disced with a heavy disc plow with two gangs of 100-cm diameter blades to a depth of about 10 cm. The disc plow was pulled by a tractor and two passes were made within each plot on each date of discing.

Vegetation Sampling

We visited each of the study areas during May–June 2003 to evaluate potential study plots and determine soil textures. Soil samples were collected with an Oakfield 36^2 Tube Sampler with Step soil probe (Oakfield Apparatus, Inc., Oakfield, Wisconsin). We overlaid a 50 × 50-m grid over the potential site and extracted 10 cm of soil at each grid point (n=18) contained within the block (100×350 m). Soil samples were analyzed at the Texas A&M University-Kingsville soil laboratory to verify soil texture. Blocks meeting the soil-texture criteria were delineated and georeferenced using a Garmin eTrex Venture Global Positioning System (Garmin International, Olathe, Kansas). Coordinates were entered into a geographic information system (GIS) using ArcView (ESRI, Redlands, California) to digitize soil-type blocks.

We collected pretreatment data on percent bare ground, forb density, visual obstruction, and non-native grass density during September 2003. We also collected post-treatment data on these four variables during three sampling periods (March, May, and July) for two years post discing (2004 and 2005). One observer collected all data to minimize observer variability. We established a 100-m transect through the center of each experimental unit. We visually estimated percent bare ground within a 20×50 -cm sampling frame (Daubenmire 1959) at 10-m intervals beginning at 10 m and terminating at 90 m (nine sampling frames/experimental unit). We also identified plants to species and recorded number of individuals for each species within each sampling frame. We used these data to estimate forb density (forbs m⁻²) and non-native grass density (non-native plants m⁻²).

We measured visual obstruction using a 1 m \times 30.5-cm vegetation profile board (Nudds 1977) that was marked in 1-dm intervals. Initial readings were taken at a distance of 4 m and a height of 0.5 m in a random direction at 10-m intervals along the transect. Three subsequent readings were obtained at each sampling point at 90°, 180°, and 270° from the first reading. We chose this distance and height because it resulted in the greatest variability in visual obstruction readings (based on pilot study) and represented the approximate height of a mammalian predator of bobwhites, respectively. Visual obstruction was averaged over strata and the four readings for each 10-m interval and then averaged for the transect.

Statistical Analysis

We tested for treatment differences using an *F* test for each soil \times time combination because of significant interactions (*P*>0.05) between treatments, soil texture, and time. We then used Tukey's

HSD mean separation test within these combinations to identify which means differed. In addition, we evaluated percent bare ground among treatments based on the bounds of habitat suitability (15%–60%) identified for bobwhites by Kopp et al. (1998).

Results

We do not discuss specific differences in percentage bare ground among treatments because it is obvious that discing will increase bare ground and that the most recent discing treatment generally will have the greatest exposure. Rather, we present data on the duration of the discing effect (i.e., when differences among treatments ceased to exist) and habitat suitability bounds.

Percentage bare ground differed (P<0.05) among treatments only during the initial part (March–May) of the first growing season in both soil textures in the Rio Grande Plains (Table 1). Percentage bare ground remained within the habitat-suitability bounds of bobwhites during this time period and approached the lower bound by July 2004 (Figures 1A and 1B). Discing had a similar treatment life in the Rolling Plains. Percentage bare ground differed (P<0.05) among treatments through the end of the first growing season in clay soils but only through March in sandy soils (Table 1). Suitable bare ground exposure for bobwhites lasted longer in the Rolling Plains, approximating the lower suitability bound during May of the second growing season. (Figures 1C and 1D).

Table 1. Percent bare ground pre- and post-treatment following four treatments (control, Oct discing, Jan discing, and Mar discing) on clay and sandy plots in the Rio Grande (Live Oak and McMullen Counties) and Rolling Plains (Hemphill County) of Texas, March–July 2003–2005. For each ecoregion and soil texture, means in a column accompanied by the same letter are not significantly (P > 0.05) different based on Tukey's test.

		20	03	2004					2005						
Soil Treatment		Pretrea	Pretreatment		ar	May		Ju	ıl	Mar		Мау		Jul	
	n	Mean	SE	Mean	SE	Mean	SE	Mean	SE	Mean	SE	Mean	SE	Mean	SE
						Rio	Grande Plai	ns Ecoregion							
Clay															
Control	6	17.2 ^B	3.1	19.3 ^B	6.2	15.7 ^B	3.6	8.0 ^A	1.2	11.7 ^A	3.6	12.4 ^A	3.1	15.8 ^A	3.2
Oct 2003	3	30.0 ^{AB}	6.0	43.3 ^{AB}	14.7	37.4 ^{AB}	10.0	21.5 ^A	5.0	10.7 ^A	3.3	24.4 ^A	5.6	16.1 ^A	4.3
Jan 2004	3	51.3 ^A	11.5	62.0 ^A	11.0	30.2 ^{AB}	6.1	13.5 ^A	6.1	8.5 ^A	6.7	20.9 ^A	18.7	13.0 ^A	7.7
Mar 2004	3	38.5 ^{AB}	9.6	70.6 ^A	29.4	51.3 ^A	11.5	14.8 ^A	3.6	17.8 ^A	4.2	22.8 ^A	11.2	23.1 ^A	8.4
Sandy															
Control	6	10.8 ^B	2.0	8.8 ^C	1.4	10.8 ^B	2.0	15.5 ^A	7.7	8.4 ^A	2.8	21.3 ^A	4.4	24.0 ^A	2.4
Oct 2003	3	10.9 ^B	1.8	5.9 ^C	2.4	15.7 ^B	5.1	32.8 ^A	2.9	4.4 ^A	2.0	13.0 ^A	2.1	32.6 ^A	9.7
Jan 2004	3	54.6 ^A	10.4	43.1 ^B	3.1	10.9 ^B	1.8	14.4 ^A	6.5	8.3 ^A	5.6	16.7 ^A	2.2	19.3 ^A	8.0
Mar 2004	3	15.7 ^B	5.1	100.0 ^A	0.0	54.6 ^A	10.4	17.4 ^A	3.4	11.5 ^A	4.6	11.7 ^A	1.5	17.0 ^A	1.3
						Ro	olling Plains	Ecoregion							
Clay															
Control	6	6.9 ^A	2.9	28.7 ^B	6.4	41.7 ^C	3.2	24.5 ^B	3.1	17.1 ^A	5.6	15.5 ^A	3.5	16.8 ^A	3.5
Oct 2003	3	7.6 ^A	5.2	42.6 ^A	6.3	48.0 ^{BC}	8.0	21.9 ^B	5.1	16.5 ^A	9.5	10.6 ^A	0.3	12.2 ^A	1.2
Jan 2004	3	8.3 ^A	3.3	68.7 ^A	5.5	71.1 ^{AB}	8.7	45.6 ^{AB}	13.8	35.4 ^A	13.1	16.7 ^A	5.0	16.7 ^A	3.9
Mar 2004	3	5.4 ^A	3.4	71.9 ^A	11.8	73.3 ^A	1.8	50.6 ^A	4.4	24.1 ^A	14.6	20.0 ^A	7.3	20.0 ^A	7.3
Sandy															
Control	6	20.8 ^A	6.8	40.2 ^A	12.7	63.1 ^A	5.4	39.2 ^A	4.3	29.6 ^A	8.1	19.5 ^A	6.2	16.8 ^A	3.4
Oct 2003	3	28.0 ^A	15.0	50.4 ^A	18.7	56.9 ^A	11.4	35.0 ^A	8.0	38.9 ^A	6.9	8.9 ^A	1.7	10.0 ^A	2.8
Jan 2004	3	8.5 ^A	3.6	59.8 ^{AB}	18.2	68.3 ^A	15.0	37.8 ^A	8.3	42.0 ^A	4.0	16.3 ^A	6.0	17.0 ^A	5.2
Mar 2004	3	9.8 ^A	7.7	78.3 ^B	9.7	62.0 ^A	12.4	34.6 ^A	4.2	36.9 ^A	14.2	9.8 ^A	1.5	12.8 ^A	2.0

Rolling Plains



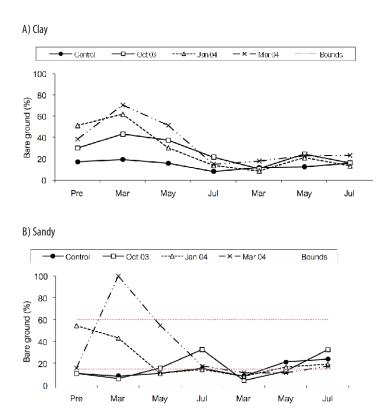
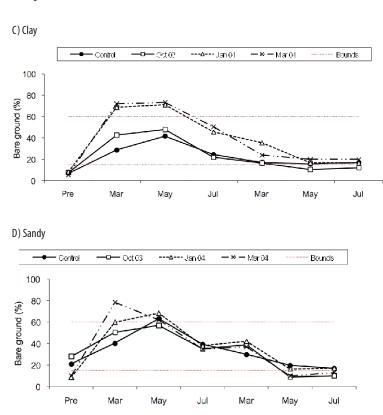


Figure 1. Bare ground exposure (%) in relation to bounds of habitat suitability for northern bobwhite among four discing treatments (control, Oct 2003, Jan 2004, and Mar 2004) on clay and sandy soils in the Rio Grande Plains (A,B) and Rolling Plains (C,D) of Texas, 2003–2005. Pretreatment data were collected on September 2003 and vegetation sampling occurred March–July 2004 and 2005.

We documented a diversity of forbs and grasses during our study (Table 2). Mean forb density (forbs m⁻²) differed among treatments during March 2004 and March 2005 in clay-textured soils in the Rio Grande Plains (Table 3). Sandy-textured soils exhibited differences in mean forb density only in March 2004. Seasonal effects appeared to have influenced fluctuations in forb density because forb density generally peaked during March in both years independent of treatment or soil texture (Figures 2A and 2B). We observed similar trends in the Rolling Plains, except these trends were expressed about one month later than in the Rio Grande Plains. Mean forb density differed among treatments during May 2004 in clay soils and May 2005 in sandy soils (Table 3). Seasonal effects also were observed in forb density in this ecoregion; forb density generally peaked during May in both years independent of treatment or soil texture (Figures 2C and 2D).



Mean visual obstruction (%) did not differ (P>0.05) among treatments throughout the study in clay soils in the Rio Grande Plains (Table 4). Mean visual obstruction differed (P<0.05) only during the first part of the growing season (March–May 2004) in sandy soils (Table 3; Figures 3A and 3B). In the Rolling Plains, mean visual obstruction differed (P<0.05) during March 2004 in clay and sandy soils (Table 3; Figures 3A and 3B). Thus, across all soil textures and in both ecoregions, mean visual obstruction was similar (P>0.05) among treatments by the end of the first growing season (Table 4).

We observed a general, positive trend for non-native grass density from pretreatment to two years post-discing in both soil types and all treatments in the Rio Grande Plains (Table 5). Non-native grass density increased (P<0.05) only in sandy soils of disced plots (Table 4). We documented no significant changes (P>0.05) in denTable 2. Common forb and grass species found on clay and sandy plots in the Rio Grande (Live Oak and McMullen counties) and Rolling Plains (Hemphill County) of Texas, March–July 2003–2005.

	Clay		Sano	ly		
Гуре	Common name	Scientific name	Common name	Scientific name		
		Rio Grande I	Region			
orb	Western ragweed	Ambrosia psilostachya	Western ragweed	Ambrosia psilostachya		
	Texas thistle	Cirsium texanum	Texas thistle	Cirsium texanum		
	Engelmann daisy	Engelmannia pinnatifida	Wooly croton	Croton capitatus		
	Common sunflower	Helianthus annuus	Heart-sepal wild buckwheat	Eriogonum multiflorum		
	Wild alfafa	Medicago sativa	Common sunflower	Helianthus annuus		
	Sour clover	Oxalis dillenii	Horsemint	Monarda citriodaro		
	Tallowweed	Plantago hookeriana	Tallowweed	Plantago hookeriana		
	Redseed plantain	Plantago rhodosperma	Upright prairie coneflower	Ratibida columnaris		
	American nightshade	Solanum ptycanthum	Silverleaf nightshade	Solanum eleagnifolium		
	Fan-leaf vervain	Verbena plicata	Fan-leaf vervain	Verbena plicata		
rass	Japanese broome	Bromus japonicus	Japanese broome	Bromus japonicus		
	Cheatgrass	Bromus tectorum	Cheatgrass	Bromus tectorum		
	Buffalograss	Buchloe dactyloides	Canadian wildrye	Elymus canadensis		
	Ryegrass	Lolium perenne	Sand lovegrass	Eragrostis trichodes		
	Common witchgrass	Panicum capillare	Ryegrass	Lolium perenne		
	Vinemesquite	Panicum obtusum	Common witchgrass	Panicum capillare		
	Switchgrass	Panicum virgatum	Vinemesquite	Panicum obtusum		
	Marsh hay cordgrass	Spartina patens	Switchgrass	Panicum virgatum		
	Alkali sacaton	Sporobolus airoides	Alkali sacaton	Sporobolus airoides		
	Six weeks grass	Vulpia octoflora	Sand dropseed	Sporobolus cryptandrus		
	j	Rolling Plains				
orb	Western ragweed	Ambrosia psilostachya	Western ragweed	Ambrosia psilostachya		
010	Arkansas lazy daisy	Aphanostephus skirrhobasis	Horsemint	Monarda citriodaro		
	Texas thistle	Cirsium texanum	Sour clover	Oxalis dillenii		
	Wild geranium	Geranium carolinianum	Wooly croton	Croton capitatus		
	Bur-clover	Medicago polymorpha	Dayflower	Commelina erecta		
	Horsemint	Monarda citriodaro	Deer-pea vetch	Vicia Iudoriciana		
	Sour clover	Oxalis dillenii	Old Man's beard	Clematis drummondii		
	Tallowweed	Plantago hookeriana	Tallowweed	Plantago hookeriana		
	Upright prairie coneflower	Ratibida columnaris	Campherweed	Pluchea camphorata		
	Deer-pea vetch	Vicia Iudoriciana	Texas thistle	Cirsium texanum		
rass	Purple threeawn	Aristida purpurea	Bermudagrass	Cynodon dactylon		
1455	King ranch bluestem	Bothriochloa ischaemum	Fescue	Festuca versuta		
	Buffelgrass	Cenchrus ciliaris	Texas wintergrass	Stipa leucotricha		
	Hooded windmill grass	Chloris cucullata	King ranch bluestem	Stipa ieucotricna Bothriochloa ischaemum		
	Bermudagrass	Cynodon dactylon	Hooded windmill grass	Bothriochioa ischaemum Chloris cucullata		
	Kleberg bluestem	Dichanthium annulatum	Buffelgrass	Chioris cucultata Cenchrus ciliaris		
	Plains lovegrass	Eragrostis intermedia	Purple threeawn	Cenchrus ciliaris Aristida purpurea		
	Hall panicum	Panicum halli	Red lovegrass	Eragrostis secundiflora		
	Johnsongrass	Sorghum halepense	Six weeks grass	Eragrostis secundifiora Vulpia octoflora		
	Texas wintergrass	Sorgnum nulepense Stipa leucotricha	Grassbur	Cenchrus incertus		

sity of non-native grasses in either clay or sandy soils in the Rolling Plains (Table 5).

Discussion

Habitat changes resulting from discing were short-lived. Differences among treatments in percentage bare ground and forb density ceased to exist by the end of the first growing season. Short-term (<2 years) improvements in habitat quality for northern bobwhites following discing also appear common in mesic ecosystems (Stoddard 1931, Buckner and Landers 1979, Jones et al. 1993, Greenfield et al. 2002). Considering that our study was conducted in more xeric rangelands, we anticipated longer-lasting treatment effects resulting from slower recovery of vegetation after disturbance. The short-lived treatment effects of discing in our study were likely the result of above-average rainfall. The study sites in the Rio Grande and Rolling Plains received about 30 cm and 20 cm of rainfall, respectively, above long-term annual means during the first year post-discing and average rainfall the year after. Because rainfall exerts a profound influence on vegetation dynamics in semi-arid rangelands, this occurrence of abundant rainfall likely shortened treatment life. Fulbright (1999, 2004) documented that habitat response to discing could be variable and inconsistent from year to year in semiarid rangelands depending on rainfall.

In our study, seasonal effects appeared to have a stronger influence on forb density than discing or soil texture. We observed consistent seasonal fluctuations in forb density in both ecoregions somewhat independent of treatment or soil texture. Forb density generally peaked in March in the Rio Grande Plains and May in Table 3. Forb density (plants m⁻²) pre- and post-treatment following four treatments (control, Oct discing, Jan discing, and Mar discing) on clay and sandy plots in the Rio Grande (Live Oak and McMullen counties) and Rolling Plains (Hemphill County) of Texas, March–July 2003–2005. For each ecoregion and soil texture, means in a column accompanied by the same letter are not significantly (P >0.05) different based on Tukey's test.

		20	03	2004						2005					
Soil		Pretreatment		Mar		May		Ju	l	Mar		Мау		Jul	
Treatment	n	Mean	SE	Mean	SE	Mean	SE	Mean	SE	Mean	SE	Mean	SE	Mean	SE
						Rio	Grande Pla	ins Ecoregion							
Clay															
Control	6	13.1 ^A	4.8	119.1 ^A	17.1	53.0 ^A	8.6	31.3 ^A	7.2	81.9 ^B	8.1	61.5 ^A	10.8	41.9 ^A	9.5
Oct 2003	3	18.5 ^A	6.8	127.4 ^A	44.0	95.9 ^A	20.5	33.3 ^A	5.7	106.7 ^{AB}	28.6	48.1 ^A	14.1	43.0 ^A	9.3
Jan 2004	3	18.9 ^A	5.3	19.3 ^B	2.6	47.0 ^A	8.2	37.8 ^A	1.9	153.7 ^A	6.3	43.3 ^A	6.3	28.9 ^A	1.3
Mar 2004	3	22.2 ^A	6.3	95.9 ^{AB}	95.9	39.3 ^A	12.3	45.6 ^A	12.4	90.7 ^B	17.3	63.3 ^A	8.0	52.6 ^A	15.5
Sandy															
Control	6	17.2 ^A	4.3	102.4 ^{AB}	31.9	80.2 ^A	20.0	27.8 ^A	7.2	74.4 ^A	21.7	24.8 ^A	6.7	25.6 ^A	2.5
Oct 2003	3	13.3 ^A	1.9	144.8 ^A	12.5	94.8 ^A	11.6	37.0 ^A	6.2	55.6 ^A	12.0	28.5 ^A	3.2	39.6 ^A	10.0
Jan 2004	3	20.7 ^A	1	78.1 ^B	15.5	115.9 ^A	18.0	61.9 ^A	17.2	53.7 ^A	14.6	46.7 ^A	10.3	33.0 ^A	5.0
Mar 2004	3	19.3 ^A	3.6	0.0 ^C	0.0	57.0 ^A	4.9	62.2 ^A	17.6	96.7 ^A	52.1	31.1 ^A	5.3	20.0 ^A	4.8
						R	olling Plain	s Ecoregion							
Clay															
Control	6	74.1 ^A	10.9	0.2 ^A	0.2	43.9 ^B	8.7	43.5 ^A	3.5	37.0 ^A	6.0	58.1 ^A	11.0	46.9 ^A	3.9
Oct 2003	3	73.7 ^A	20.2	0.0 ^A	0.0	103.7 ^A	24.3	45.2 ^A	7.8	59.6 ^A	2.4	84.1 ^A	35.3	38.9 ^A	5.9
Jan 2004	3	63.7 ^A	25.7	0.0 ^A	0.0	42.2 ^B	8.4	20.7 ^A	7.7	39.6 ^A	11.2	75.6 ^A	18.4	40.0 ^A	1.3
Mar 2004	3	81.1 ^A	25.5	0.0 ^A	0.0	67.0 ^A	2.3	53.7 ^A	3.5	29.6 ^A	1.5	71.9 ^A	14.3	50.4 ^A	8.4
Sandy															
Control	6	15.9 ^A	3.6	0.2 ^A	0.2	24.6 ^A	5.8	18.3 ^A	2.9	29.6 ^A	3.7	68.5 ^A	6.6	40.0 ^A	13.9
Oct 2003	3	11.1 ^A	4.5	0.0 ^A	0.0	79.6 ^A	46.3	55.2 ^A	18.9	35.6 ^A	9.7	25.2 ^B	10.9	19.3 ^A	7.5
Jan 2004	3	8.9 ^A	5	0.0 ^A	0.0	46.3 ^A	31.9	45.2 ^A	7.0	40.0 ^A	2.3	47.0 ^{AB}	22.3	20.0 ^A	7.8
Mar 2004	3	12.6 ^A	2.7	0.0 ^A	0.0	69.6 ^A	21.5	38.1 ^A	3.6	58.5 ^A	21.9	37.0 ^B	9.1	31.5 ^A	2.3

Table 4. Mean visual obstruction (%) pre- and post-treatment following four treatments (control, Oct discing, Jan discing, and Mar discing) on clay and sandy plots in the Rio Grande (Live Oak and McMullen counties) and Rolling Plains (Hemphill County) of Texas, March–July 2003–2005. For each ecoregion and soil texture, means in a column accompanied by the same letter are not significantly (P >0.05) different based on Tukey's test.

		200)3	2004						2005					
Soil		Pretreatment		Mar		May		Ju	I	Mar		May		Jul	
Treatment	n	Mean	SE	Mean	SE	Mean	SE	Mean	SE	Mean	SE	Mean	SE	Mean	SE
						Rio G	irande Plai	ns Ecoregion							
Clay															
Control	6	60.3 ^A	7.3	7.5 ^A	1.3	47.8 ^A	3.1	71.2 ^A	2.3	44.5 ^A	5.0	49.1 ^A	4.4	40.3 ^A	4.0
Oct 2003	3	48.4 ^A	10.6	2.2 ^A	0.5	34.6 ^A	4.5	54.8 ^A	4.1	27.6 ^A	4.5	35.8 ^A	2.1	36.7 ^A	3.9
Jan 2003	3	68.8 ^A	6.1	0.8 ^A	0.2	37.8 ^A	4.5	60.2 ^A	5.8	35.2 ^A	4.6	30.1 ^A	5.2	39.6 ^A	6.0
Mar 2004	3	70.7 ^A	6.7	3.3 ^A	1.7	20.7 ^A	2.5	83.4 ^A	7.7	48.2 ^A	9.1	38.9 ^A	7.3	32.8 ^A	2.1
Sandy															
Control	6	78.9 ^A	3.6	48.6 ^A	5.5	77.5 ^{AB}	4.5	79.9 ^A	3.2	42.5 ^A	6.6	84.5 ^A	5.1	87.7 ^A	3.2
Oct 2003	3	70.3 ^A	5.5	35.0 ^A	5.5	95.9 ^A	2.0	90.5 ^A	3.6	61.1 ^A	5.9	62.9 ^A	9.3	99.1 ^A	0.6
Jan 2003	3	86.6 ^A	3.9	2.5 ^B	0.2	56.1 ^B	3.9	90.2 ^A	3.7	41.8 ^A	9.5	79.8 ^A	7.6	98.8 ^A	1.3
Mar 2004	3	87.2 ^A	3.4	0.0 ^B	0.0	24.7 ^C	5.2	90.4 ^A	3.2	29.1 ^A	7.9	74.1 ^A	8.7	89.0 ^A	3.8
						Ro	lling Plains	Ecoregion							
Clay															
Control	6	83.6 ^A	5.5	52.1 ^A	5.4	17.6 ^A	2.1	37.3 ^A	3.7	28.1 ^A	1.9	31.2 ^A	1.1	68.4 ^A	4.5
Oct 2003	3	98.3 ^A	1.1	0.0 ^B	0.0	4.5 ^A	0.4	33.1 ^A	3.0	27.6 ^A	1.5	31.4 ^A	2.2	71.2 ^A	7.6
Jan 2003	3	81.6 ^A	8.9	0.0 ^B	0.0	3.5 ^A	0.4	29.3 ^A	4.3	25.3 ^A	3.3	30.9 ^A	1.7	71.1 ^A	8.1
Mar 2004	3	86.2 ^A	5.6	0.0 ^B	0.0	3.8 ^A	0.4	48.4 ^A	4.5	30.0 ^A	0.9	26.3 ^A	1.6	75.2 ^A	5.6
Sandy															
Control	6	72.8 ^A	5.7	70.6 ^A	7.6	22.7 ^A	3.6	70.2 ^A	5.1	54.6 ^A	4.5	37.6 ^A	1.3	64.7 ^A	4.8
Oct 2003	3	54.1 ^{AB}	10.4	0.0 ^B	0.0	6.8 ^A	0.9	75.5 ^A	6.1	32.8 ^A	4.8	34.6 ^A	0.6	64.4 ^A	5.1
Jan 2003	3	76.1 ^A	10.3	0.0 ^B	0.0	8.5 ^A	1.2	84.2 ^A	6.9	57.4 ^A	3.1	36.9 ^A	0.6	75.9 ^A	6.5
Mar 2004	3	35.9 ^B	4.0	0.0 ^B	0.0	8.3 ^A	1.2	86.2 ^A	6.2	48.1 ^A	6.1	34.4 ^A	0.7	73.7 ^A	5.3

Rio Grande Plains

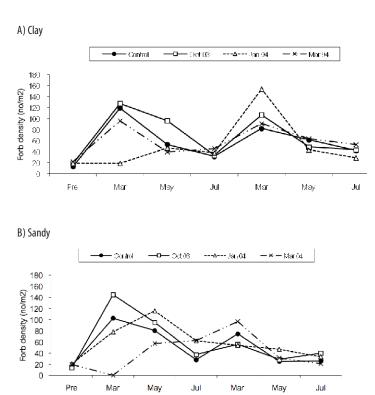


Figure 2. Forb density (plants m⁻²) among four discing treatments (control, Oct 2003, Jan 2004, and Mar 2004) on clay and sandy soils in the Rio Grande Plains (A,B) and Rolling Plains (C,D) of Texas, 2003–2005. Pretreatment data were collected on September 2003 and sampling occurred March–July 2004 and 2005.

the Rolling Plains. Peak forb density in spring in both ecoregions resulted from response of cool-season species. The one-month delay in peak forb density in the Rolling Plains was expected because the higher latitude of this site resulted in a relatively longer and harsher winter season.

An interesting finding of our study was that discing appeared to accelerate establishment of non-native grasses in the Rio Grande Plains. Although density of non-native grasses increased on both control and disced plots, percent increase of these grasses was greater numerically on disced plots, particularly those in sandy soils. We did not observe such a pronounced response of non-native grasses in the Rolling Plains. These contrasting results may be related to ecological and physiological characteristics of the nonnative species involved. Kleberg bluestem, King Ranch bluestem, and bermudagrass were the non-native species found in the Rio Grande Plains study sites. These grasses are warm-season, peren**Rolling Plains**

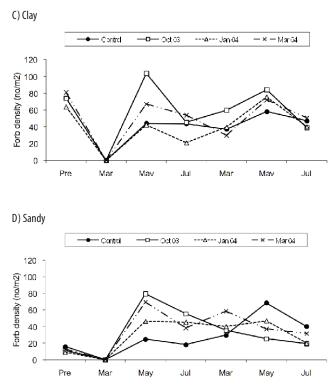
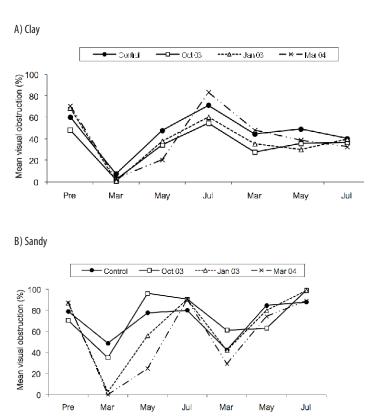


Table 5. Percent change in density of non-native grasses (plants m⁻²) from pre-discing (Sep 2003) to two-years post-discing (Jul 2005) in the Rio Grande (Live Oak and McMullen counties) and Rolling Plains (Hemphill County) of Texas.

Soil		200 Pretreat	-	200 Ju		Differ	ence		
Treatment	n	Mean	SE	Mean	SE	Mean	SE	% Change	P-value
			Rio	Grande	Plains I	Ecoregion			
Clay									
Control	6	23.9	7.8	38.9	16.4	15.0	11.2	62.8%	0.24
Disced	9	13.7	6.9	28.0	14.4	14.3	7.8	104.4%	0.10
Sandy									
Control	6	7.1	2.5	33.3	14.6	26.3	14.8	369.0%	0.14
Disced	9	10.0	2.1	54.2	14.9	44.2	14.0	442.0%	0.01
			F	Rolling Pl	ains Eco	oregion			
Clay									
Control	6	8.9	4.0	13.9	3.9	5.0	3.1	56.2%	0.17
Disced	9	3.5	1.0	8.2	2.2	4.7	2.4	134.3%	0.08
Sandy									
Control	6	35.9	7.1	27.1	4.7	-8.9	4.2	-24.5%	0.90
Disced	9	41.9	5.8	29.5	8.5	-12.4	11.4	-29.6%	0.31

Rio Grande Plains



Rolling Plains

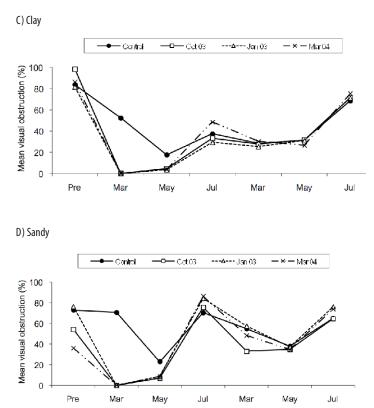


Figure 3. Mean visual obstruction (%) among four discing treatments (control, Oct 2003, Jan 2004, and Mar 2004) on clay and sandy soils in the Rio Grande Plains (A,B) and Rolling Plains (C,D) of Texas, 2003–2005. Pretreatment data were collected on September 2003 and sampling occurred March–July 2004 and 2005.

nial grasses that photosynthesize by the C_4 pathway. These C_4 species complete most of their growth during warmer portions of the year after the C_3 plants have completed their growth. The invasive grass species in the Rolling Plains study sites, cheatgrass, Japanese brome, and perennial ryegrass, are C_3 species that complete most of their growth at the same time as the period of maximum growth of cool season forbs. Additionally, cheatgrass and Japanese brome are both annuals that senesce as seeds mature during the summer. Apparent differences in the response of non-native grass species between the Rio Grande Plains and Rolling Plains may have resulted in part because of differences in season of growth relative to the timing of discing and longevity of the non-native grass species present.

Our findings are consistent with others reporting a facilitating, but variable role of soil disturbance in establishment and spread of non-native plants (Brown and Peet 2003, D' Antonio et al. 1999). Studies have reported mixed responses of invasive species to disturbance ranging from increasing (Johnson and Fulbright 2008) to decreasing (Madison et al. 2001) to no-effect (Hobbs and Atkins 1988). These mixed responses may be influenced by soil physical and chemical properties and non-native plant species involved. Johnson and Fulbright (2008) reported canopy cover of buffelgrass (*Pennisetum ciliare*) was not affected by discing on Delmita soils, but exhibited a 10-fold increase five-years post discing on Ramadero soils. The possible role discing may have in facilitating establishment of non-native grasses is of concern because non-native grasses generally do not represent suitable bobwhite habitat (Kuvlesky et al. 2002, Sands 2006). Flanders et al. (2006) reported sites dominated by non-native plants had bobwhite densities about 50% lower than sites dominated by native plants.

Management Implications

Timing of discing did not appear to have a profound impact on habitat structure or duration of treatment life in our study. Our findings indicate the structural effects of discing on bobwhite habitat can be short-lived (<1.5 years) on semi-arid rangelands, and maintaining bobwhite habitat through discing is a biennial management practice. A management concern is density of non-native grasses may increase following discing, particularly when invasive species are involved. This finding is important because non-native grasses generally do not represent suitable bobwhite habitat. In making decisions regarding habitat manipulation, managers must weigh the temporary benefits derived from discing against the potential long-term detrimental impacts on bobwhite habitat resulting from increased density of non-native grasse

Acknowledgments

We thank the Caesar Kleberg Wildlife Research Institute and Texas A&M University for providing financial and logistical support. Cooperative funding was provided by the Texas Parks and Wildlife Department; Texas State Council, South Texas, Houston, East Texas, and Alamo chapters of Quail Unlimited; George and Mary Josephine Hamman Foundation; Robert J. Kleberg, Jr. and Helen C. Kleberg Foundation; Amy Shelton McNutt Charitable Trust; Amy Shelton McNutt Memorial Fund; Bob and Vivian Smith Foundation; and The William A. and Madeline Welder Smith Foundation. We thank William P. Kuvlesky, Jr., and Alfonso Ortega-Sanchez for providing helpful comments on earlier versions of this manuscript. This manuscript is Caesar Kleberg Wildlife Research Institute publication number 09–103.

Literature Cited

- Bailey, J. A. 1984. Principles of wildlife management. John Wiley and Sons, New York, New York.
- Brown, R. L. and R. K. Peet. 2003. Diversity and invasibility of southern Appalachian plant communities. Ecology 84:32–39.
- Buckner, J. L. and J. L. Landers. 1979. Fire and discing effects on herbaceous food plants and seed supplies. Journal of Wildlife Management 43:807-811.
- D'Antonio, C. M., T. L. Dudley, and M. Mack. 1999. Disturbance and biological invasions: direct effects and feedbacks. Pages 413–452 in L. R. Walker, editor. Ecosystems of the World 16: Ecosystems of disturbed ground. Elsevier Science, Amsterdam, Netherlands.
- Daubenmire, R. 1959. A canopy coverage method of vegetation analysis. Northwest Science 33:43–65.
- Flanders, A. A., W. P. Kuvlesky, Jr., D. C. Ruthven III, R. E. Zaiglen, R. L. Bingham, T. E. Fulbright, F. Hernandez, and L. A. Brennan. 2006. Effects of invasive exotic grasses on South Texas rangeland breeding birds. Auk 123:171–182.
- Fulbright, T. E. 1999. Response of white-tailed deer foods to discing in a semiarid habitat. Journal of Range Management 52:346–350.

— 2004. Disturbance effects on species richness of herbaceous plants in a semi-arid habitat. Journal of Arid Environments 58:119–133.

- Greenfield, K. C., L. W. Burger, M. J. Chamberlain, and E. W. Kurzejeski. 2002. Vegetation management practices on Conservation Reserve Program fields to improve northern bobwhite habitat quality. Wildlife Society Bulletin 30:527– 538.
- Guthery, F. S. 1986. Beef, brush, and bobwhites: quail management in cattle country. Caesar Kleberg Wildlife Research Institute Press, Texas A&I University, Kingsville.
- Gould, F. W. 1975. Texas plants: a checklist and ecological summary. Texas A&M University, Agricultural Experiment Station, College Station.
- Hobbs, R. J. and L. Atkins. 1988. Effect of disturbance and nutrient addition on native and introduced annuals in plant communities in the Western Australian wheatbelt. Australian Journal of Ecology 13:171–179.
- Jackson, A. S. 1972. Quail Management Handbook. Texas Parks and Wildlife Department, Austin, Bulletin No. 48.
- Jackson, J. R., G. A. Hurst, and E. A. Gluesing. 1987. Abundance and selection of invertebrates by northern bobwhite chicks. Proceedings of the Annual Conference of the Southeastern Association of Fish and Wildlife Agencies 41:303– 310.
- Johnson, M. V. and T. E. Fulbright. 2008. Is exotic plant invasion enhanced by a traditional wildlife habitat management technique? Journal of Arid Environments 72:1911–1917.
- Jones, P. D., J. R. Sweeney, and T. Ivey. 1993. Effects of six discing regimes on quail foods in fallow fields. Proceedings of the Annual Conference of the Southeastern Association of Fish and Wildlife Agencies 47:239–250.
- Kopp, S. D., F. S. Guthery, N. D. Forrester, and W. E. Cohen. 1998. Habitat selection modeling for northern bobwhites on subtropical rangeland. Journal of Wildlife Management 62:884–894.
- Kuvlesky, W. P., Jr., T. E. Fulbright, and R. Engel-Wilson. 2002. The impact of exotic grasses on quail in the southwestern United States. Proceedings of the National Quail Symposium 5: 118–128.
- Lehmann, V. M. 1984. Bobwhites in the Rio Grande Plains of Texas. Texas A&M University Press, College Station.
- Madison, L. A., T. G. Barnes, and J. D. Sole. 2001. Effectiveness of five discing and herbicide to renovate tall fescue fields to northern bobwhite habitat. Wildlife Society Bulletin 29:706–712.
- Manley, S. W., R. S. Fuller, J. M. Lee, and L. A. Brennan. 1994. Arthropod response to strip discing in old fields managed for northern bobwhites. Proceedings of the Annual Conference of the Southeastern Association of Fish and Wildlife Agencies 48:227–235.
- Nudds, D. 1977. Quantifying the vegetative structure of wildlife cover. Wildlife Society Bulletin 5:113–117.
- Patton, D. R. 1992. Wildlife habitat relationships in forested ecosystems. Timber Press, Portland, Oregon.
- Peoples, A. D., R. L. Lochmiller, D. M. Leslie, Jr., and D. M. Engle. 1994. Producing northern bobwhite food on sandy soils in semiarid mixed prairies. Wildlife Society Bulletin 22:204–211.
- Robinson, W. L. and E. G. Bolen. 2003. Wildlife ecology and management. Macmillan Publishing Company, New York, New York.
- Rosene, W. 1969. The bobwhite quail: its life and management. Rutgers University Press, New Brunswick, New Jersey.
- Sands, J. P. 2006. Impacts of invasive exotic grasses on northern bobwhite habitat use in south Texas. Thesis. Texas A&M University-Kingsville, Kingsville.
- Shelton, M. D. and C. R. Edwards. 1983. Effects of weeds on the diversity and abundance of insects in natural habitats. Journal of Animal Ecology 38:497–509.
- Spears, G. S., F. S. Guthery, S. M. Rice, S. J. DeMaso, B. Zaiglin. 1993. Optimum seral stage for northern bobwhites as influenced by site productivity. Journal of Wildlife Management 57:805–811.
- Stoddard, H. L. 1931. The bobwhite quail: its habitats, preservation, and increase. Charles Scribner's Sons, New York, New York.
- Webb, W. M. and F. S. Guthery. 1983. Response of wildlife food plants to spring discing of mesquite rangeland in northwest Texas. Journal of Range Management 36:351–353.